



ATP Biotech Inc.

**ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria)
Catalog No. ARB050/ARB100**

**ATP™ Total RNA Mini Kit (Tissue)
Catalog No. ART050/ART100**

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ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria)

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ATP™ Total RNA Mini Kit (Tissue)

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Introduction

Format : Spin column
 Sample : Up to 300 µl of whole blood
 Up to 10⁶ animal cultured cells
 Up to 10⁸ bacterial cultured cells
 Operation : Centrifuge / vacuum manifold
 Operation time : 20 minutes
 Yield : Up to 30 µg
 Application : RT-PCR、Real-Time PCR、Northern Blotting、mRNA Selection、cDNA Synthesis、Primer Extension

ATP™ RNA Mini Kit (Blood/Culture cell) is specially designed for purification of total RNA from fresh whole human blood and cultured cells. This method uses detergents and a chaotropic salt to lyse cells and inactivate RNase, and then RNA in chaotropic salt is bonded to the glass fiber matrix of the column. After washing off the contaminants, the purified RNA is eluted by RNase-free water. The entire procedure can be completed in 20 minutes and the purified RNA is ready for RT-PCR, Northern blotting, primer extension and cDNA library construction.

Kit Contents : Cat.No. / Kit Contents

| ARB50 (50 preps/kit) | ARB100 (100 preps/kit) |
|--|---|
| RBC lysis Buffer : 100 ml | RBC lysis Buffer : 200 ml |
| RT Buffer : 15 ml | RT Buffer : 30 ml |
| RB Buffer : 30 ml | RB Buffer : 60 ml |
| W1 Buffer : 30 ml | W1 Buffer : 50 ml |
| Wash Buffer (concentrated)* : 12.5 ml | Wash Buffer (concentrated)** : 25 ml |
| RNase-free water : 6 ml | RNase-free water : 6 ml |
| Filter Column : 50 pcs | Filter Column : 100 pcs |
| RB Column : 50 pcs (yellow/white filter) | RB Column : 100 pcs (yellow/white filter) |
| 2 ml Collection Tubes : 100 pcs | 2 ml Collection Tubes : 200 pcs |
| * Add 50 ml ethanol (96-100%) to Wash Buffer prior to initial use. | |
| ** Add 100 ml ethanol (96-100%) to Wash Buffer prior to initial use. | |

Cautions : RB Buffer contain guanidine hydrochloride which harmful and irritant agent.
During operation, always wear a lab coat, disposable gloves, and protective goggles.
For more information, please refer to the appropriate material safety data sheets (MSDS).

Use limitation : For research use only; not for diagnostic or medical purposes

Recommendations

1. For larger volumes of whole blood or cultured cells, ATP™ Total RNA Maxi Kits (Blood/Culture cell) is recommended.
2. For High-throughput sample processing, ATP™ 96-Well Total RNA Kit is recommended.
3. For purification of viral RNA, the ATP™ Nucleic Acid Extraction Kit is recommended. All buffers and components of this kit are guaranteed to be RNase free.

Equipments and Reagents are provided by User

- 1.5 ml microcentrifuge tubes (RNase-free)
- Microcentrifuge with rotor for 2 ml tubes
- β -Mercaptoethanol (β -ME ; 10 μl of β -ME per 1ml RB Buffer)
- PBS (phosphate-buffered saline) may be required for some samples
- Ethanol (96-100%)
- Ethanol (70%)
- DNase I (2 KU/ml, RNase-free) in reaction buffer (1 M NaCl ; 20mM Tris-HCl ; 10mM MnCl₂ ; pH 7.0 at 25°C).
- Lysozyme Buffer (20 mg/ml Lysoyme ; 20 mM Tris-HCl ; 2 mM EDTA ; 1% Triton X-100 , pH 8.0) for Gram-negative bacteria prelysis. Prepare the Lysozyme buffer fresh just before use.

Notice

We recommend to apply highly purified DNase I (RNase-free) for sensitive downstream applications. If genomic DNA contamination may be negligible or inconsequential to the application, DNase I may not be necessary to apply.

Total RNA Mini (Blood/Culture Cell) Protocol

- ARB050 : Add 50 ml ethanol (96-100%) to Wash Buffer prior to the initial use.
ARB100 : Add 100 ml ethanol (96-100%) to Wash Buffer prior to the initial use.
- Add 10 μl of β -Mercaptoethanol per 1ml of RB Buffer to RB Buffer before use.
- Additionally required : Microcentrifuge tube 、 70% Ethanol 、 DNase I (2 KU/ml, RNase-free) in reaction buffer (1 M NaCl ; 20 mM Tris-HCl ; 10 mM MnCl₂ ; pH 7.0 at 25 °C).

RBC Lysis / Cell Harvesting

Use fresh human blood:

1. Collected fresh blood in EDTA-NA₂-treated collection tubes (or other anti-coagulant mixtures).
2. Add 1 ml RBC Lysis Buffer into a sterile 1.5 ml reaction tube, and then add 300 μl human whole blood and mix by inversion. Do not vortex. Incubate the tube at room temperature for 5 min.
3. Centrifuge at 2500 rpm (500 xg) for 2 min and discard the supernatant, then add 100 μl RBC Lysis Buffer to resuspend the cell pellet.

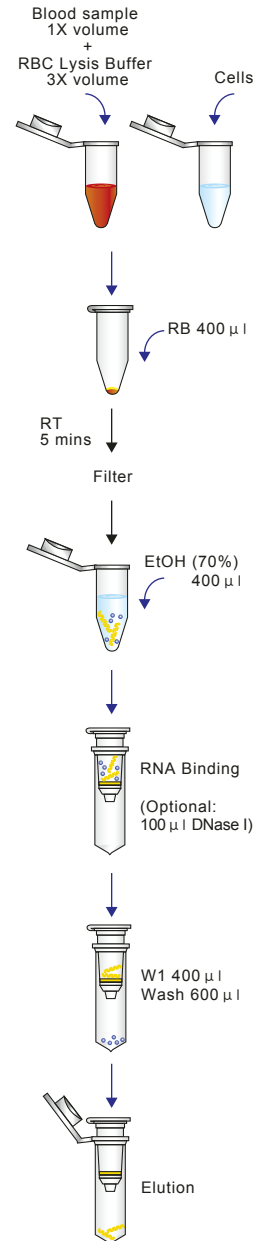
Use culture cell:

1. If using adherent cells, trypsinize the cells before harvesting.
2. Transfer 10⁶-10⁷ of cells into a microcentrifuge tube (provided by user) and harvest the cells with centrifugation for 20 seconds at 6,000 x g (8,000 rpm).
3. Discard the supernatant and resuspend the cells in 100 μl PBS or RBC Lysis Buffer.

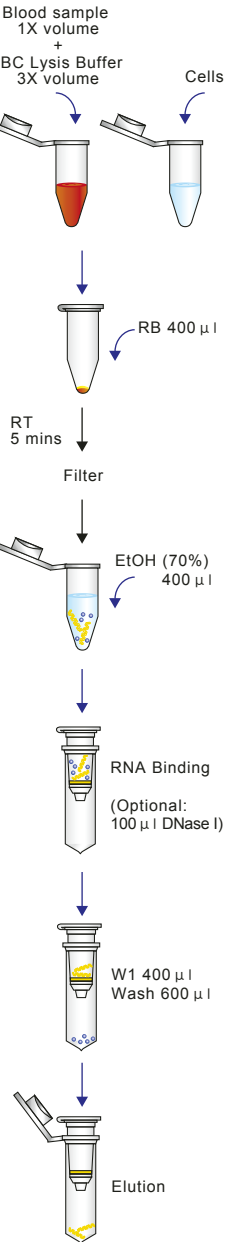
Cell Lysis

4. Add 400 μl of RB Buffer to the tube and mix with white pellet (from step 3) by vortexing.
5. Incubate at room temperature for 5 minutes
6. Place a Filter column in a 2 ml Collection tube. Apply sample mixture into the column.
7. Centrifuge at full speed (13,000 rpm) for 2 minutes.
8. Discard the Filter column and transfer the clarified filtrate in a new microcentrifuge tube (provided by user).

ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria) Blood / Culture Cell Protocol



ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria)
Blood / Culture Cell Protocol



RNA Binding

9. Place a RB Column in a 2 ml Collection Tube.
10. Add 400 µl of 70% ethanol to the sample lysate from step 8 and mix immediately by pipetting.
11. Apply 500 µl of ethanol-added mixture from previous step into the RB column.
12. Centrifuge at full speed (13,000 rpm) for 2 minute.
13. Discard the flow-through and apply the rest into the same Column.
14. Centrifuge at full speed (13,000 rpm) for 2 minute.
15. Discard the flow-through and place the RB Column back in the Collection Tube.

Optional Step: DNA residue degradation

- Add 100 µl DNase I (2 U/µl, provided by user) onto the center of the RB column matrix. Incubate for 10 minutes at room temperature.
- Go to step 16 for Washing procedure

Washing

16. Add 400 µl of W1 Buffer into the RB column. Centrifuge at 10,000 xg (13,000 rpm) for 30 seconds.
17. Discard the flow-through and place the RB Column back in the Collection Tube.
18. Add 600 µl of Wash Buffer (ethanol added) in the RB column.
19. Centrifuge at 10,000x g (13,000 rpm) for 30 seconds.
20. Discard the flow-through and return the RB Column in the 2ml Collection Tube.
21. Centrifuge again for 3 minutes at full speed to dry the column matrix.

RNA Elution

22. Transfer dried RB Column in a clean 1.5ml microcentrifuge tube (RNase-free, provided by user).
23. Add 50 µl of preheated RNase-free water into the center of the column matrix.
24. Stand for 3-5 minutes until water has been absorbed by the matrix.
25. Centrifuge at 13,000 rpm for 1 minute to elute purified RNA.

Total RNA Mini (Bacteria) Protocol

- ARB050 : Add 50 ml ethanol (96-100%) to Wash Buffer prior to the initial use. ARB100 : Add 100 ml ethanol (96-100%) to Wash Buffer prior to the initial use. Add 10 µl of β-Mercaptoethanol per 1ml of RB Buffer to RB Buffer before use.
- Additionally required : Microcentrifuge tube ∨ 70% Ethanol ∨ DNase I (2 KU/ml, RNase-free) in reaction buffer (1 M NaCl ; 20 mM Tris-HCl ; 10 mM MnCl₂ ; pH 7.0 at 25 °C).
- For Gram-negative bacteria : RT buffer
For Gram-positive bacteria : Lysozyme Buffer (20 mg/ml lysozyme ; 20 mM Tris-HCl ; 2 mM EDTA ; 1% Triton X-100 , pH 8.0). Prepare the lysozyme buffer fresh just before use.

Cell harvest

For Gram-negative bacteria

1. Transfer bacterial culture (< 10⁹) into a microcentrifuge tube (provided by user).
2. Centrifuge for 1 min at full speed (13,000 rpm) in a microcentrifuge and discard the supernatant, and then vortexing the cell pellet for 30 seconds.
3. Add 200 µl of RT Buffer to the tube and resuspend the cell pellet by vortexing or pipetting.
4. Incubate at room temperature for 5 minutes.

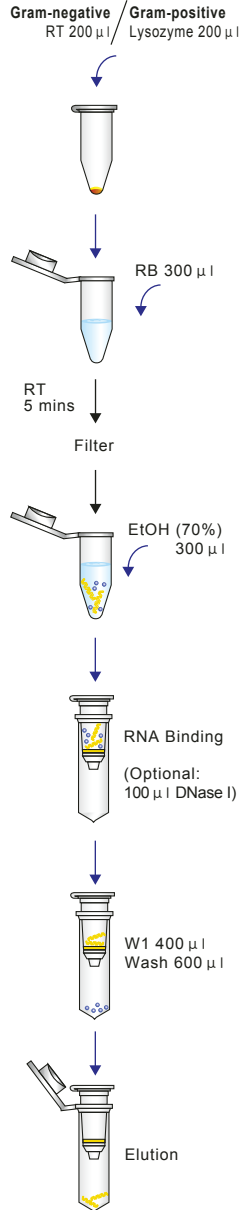
For Gram-positive bacteria

1. Transfer bacterial culture (< 10⁹) into a microcentrifuge tube (provided by user).
2. Centrifuge at full speed (13,000 rpm) for 1 min and discard the supernatant.
3. Add 200 µl of Lysozyme Buffer into the tube and resuspend the cell pellet by vortexing or pipetting.
4. Incubate at room temperature for 10 minutes. During incubation, invert the tube every 2-3min.

Cell Lysis

5. Add 300 µl RB Buffer to the tube and mix with sample lysate (from step 4) by vortexing.
6. Incubate at room temperature for 5 minutes
7. Centrifuge at full speed (13,000 rpm) for 2 minutes.
8. Transfer the supernatant into a new microcentrifuge tube.

ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria)
Bacterial Protocol





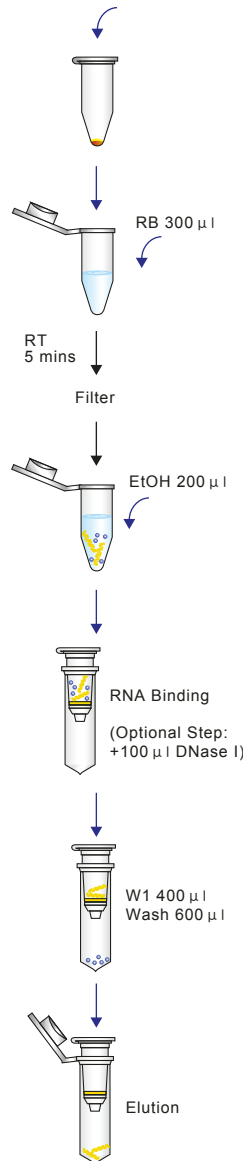
ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria)

Store at room temperature (15-25°C)

ATP™ Total RNA Mini Kit (Blood/Culture Cell/Bacteria)

Bacterial Protocol

Gram-negative RT 200 μl / Gram-positive Lysozyme 200 μl



RNA Binding

- Place a RB Column in a 2 ml Collection Tube.
- Add 300 μl of 70% ethanol to the sample lysate from step 8 and mix immediately by pipetting.
- Apply 500 μl of ethanol-added mixture from previous step into the RB column.
- Centrifuge at full speed (13,000 rpm) for 2 minute.
- Discard the flow-through and apply the rest into the same Column.
- Centrifuge at full speed (13,000 rpm) for 2 minute.
- Discard the flow-through and place the RB Column back in the Collection Tube.

Optional Step: DNA residue degradation

- Add 100 μl DNase I (2 U/μl, provided by user) onto the center of the RB column matrix. Incubate for 10 minutes at room temperature.
- Go to step 16 for Washing procedure

Washing

- Add 400 μl of W1 Buffer into the RB column. Centrifuge at 10,000 xg (13,000 rpm) for 30 seconds.
- Discard the flow-through and place the RB Column back in the Collection Tube.
- Add 600 μl of Wash Buffer (ethanol added) in the RB column.
- Centrifuge at 10,000x g (13,000 rpm) for 30 seconds.
- Discard the flow-through and return the RB Column in the 2ml Collection Tube.
- Centrifuge again for 3 minutes at full speed to dry the column matrix.

RNA Elution

- Transfer dried RB Column in a clean 1.5ml microcentrifuge tube (RNase-free, provided by user).
- Add 50 μl of preheated RNase-free water into the center of the column matrix.
- Stand for 3-5 minutes until water has been absorbed by the matrix.
- Centrifuge at 13,000 rpm for 1 minute to elute purified RNA.

ATP™ Total RNA Mini Kit (Tissue)

Store at room temperature (15-25°C)

Introduction

Format : Spin column
 Sample : Up to 25 mg of tissue
 Operation : Centrifuge / vacuum manifold
 Recovery : Up to 25 μg
 Elution volume : 50 μl
 Application : RT-PCR, Real-Time PCR, Northern Blotting, mRNA Selection, cDNA Synthesis, Primer Extension

ATP™ Total RNA Mini Kit (Tissue) is specially designed for purification of total RNA from a variety of animal tissues or cells. The provided micropestle can efficiently homogenize tissue samples in the microcentrifuge tube. The method uses detergent and a chaotropic salt to lyse cells and inactivate RNase. Then RNA in the chaotropic salt is bonded to the glass fiber matrix of the column. After washing off the contaminants, the purified RNA is eluted by RNase-free water. The entire procedure can be completed in 20 minutes and the purified RNA is ready for RT-PCR, Northern blotting, primer extension and cDNA library construction.

Kit Contents : Cat.No. / Kit Contents

| ART50 (50 prep/kit) | | ART100 (100 prep/kit) | |
|--|-----------|--|---------|
| RB Buffer | : 30 ml | RB Buffer | : 60 ml |
| W1 Buffer | : 30 ml | W1 Buffer | : 50 ml |
| Wash Buffer (concentrated)* | : 12.5 ml | Wash Buffer (concentrated)* | : 25 ml |
| RNase-free water | : 6 ml | RNase-free water | : 6 ml |
| RB Columns : 50 pcs (yellow/white filter) | | RB Columns : 100 pcs (yellow/white filter) | |
| Filter Column : 50 pcs | | Filter Column : 100 pcs | |
| 2 ml Collection Tubes : 150 pcs | | 2 ml Collection Tubes : 300 pcs | |
| Micropestle : 50 pcs | | Micropestle : 100 pcs | |
| * Add 50 ml ethanol (96-100%) to Wash Buffer prior to initial use. | | ** Add 100 ml ethanol (96-100%) to Wash Buffer prior to initial use. | |

Caution : RB Buffer contains irritant agent. During operation, always wear a lab coat, disposable gloves, and protective goggles.

Use limitation : For research use only; not for diagnostic or medical purposes

Recommendations

- For High-throughput sample processing, ATP™ 96-Well Genomic DNA Kit is recommended.
 - For purification of viral RNA, the ATP™ Nucleic Acid Extraction Kit is recommended.
- All buffers and components of this kit are guaranteed to be RNase free.

Equipments and Reagents are provided by User

- 1.5 ml microcentrifuge tubes (RNase-free)
- Microcentrifuge with rotor for 2 ml tubes
- β-Mercaptoethanol (β-ME ; 10 μl of β-ME per 1ml RB Buffer)
- 20-G needle syringe
- Ethanol (96-100%)
- Ethanol (70%)
- DNase I (2 KU/ml, RNase-free) in reaction buffer (1 M NaCl ; 20 mM Tris-HCl ; 10 mM MnCl₂ ; pH 7.0 at 25°C).

Total RNA Mini (Tissue) Protocol

- ARB050 : Add 50 ml ethanol (96-100%) to Wash Buffer prior to the initial use.
- ARB100 : Add 100 ml ethanol (96-100%) to Wash Buffer prior to the initial use.
- Add 10 μl of β-Mercaptoethanol per 1ml of RB Buffer to RB Buffer before use.
- Additionally required : Microcentrifuge tube 、 70% Ethanol 、 DNase I (2 KU/ml, RNase-free) in reaction buffer (1 M NaCl ; 20 mM Tris-HCl ; 10 mM MnCl₂ ; pH 7.0 at 25 °C).

Cell Lysis

1. Cut up to 10 mg of fresh or frozen animal tissue and transfer it into a microcentrifuge tube (provided by user).
2. Add 400 μl RB Buffer into the tube and use provided Micropestle to grind the tissue a few times.
3. Shear the tissue by passing lysate through a 20-G needle syringe 10 times.
4. Incubate at room temperature for 5 minutes.
5. Place a Filter Column in a 2 ml Collection Tube. Apply sample mixture into the column.
6. Centrifuge at full speed (13,000 rpm) for 2 minutes.
7. Discard the filter column and transfer the clarified filtrate into a new microcentrifuge tube (provided by user).

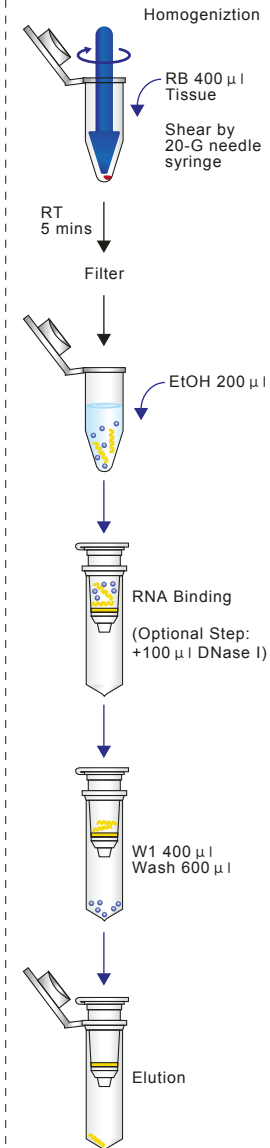
RNA Binding

8. Place a RB Column in a 2 ml Collection Tube.
9. Add 400 μl of 70 % ethanol to the sample lysate from step 7 and mix immediately by pipetting.
10. Centrifuge at full speed (13,000 rpm) for 2 minute.
11. Discard the flow-through and place the RB Column back in the Collection Tube.

Optional Step: DNA residue degradation

- Add 100 μl DNase I (2 U/μl, provided by user) onto the center of the RB column matrix. Incubate for 10 minutes at room temperature.
- Go to step 12 for Washing procedure

ATP™ Total RNA (Tissue)



ATP™ Total RNA Mini Kit (Tissue)

Store at room temperature (15-25°C)

ATP™ Total RNA (Tissue)

Washing

12. Add 400 μ l of W1 Buffer into the RB column.
13. Centrifuge at 10,000x g (13,000 rpm) for 30 seconds.
14. Discard the flow-through and place the RB Column back in the Collection Tube.
15. Add 600 μ l of Wash Buffer (ethanol added) in the RB column.
16. Centrifuge at 10,000x g (13,000 rpm) for 30 seconds.
17. Discard the flow-through and return the RB Column in the 2ml Collection Tube.
18. Centrifuge again for 3 minutes at full speed to dry the column matrix.

RNA Elution

19. Transfer dried RB Column in a clean 1.5ml microcentrifuge tube (RNase-free, provided by user).
20. Add 50 μ l of RNase-free water into the center of the column matrix.
21. Stand for 3 minutes until Elution Buffer has been absorbed by the matrix.
22. Centrifuge at 13,000 rpm for 1 minute to elute purified RNA.

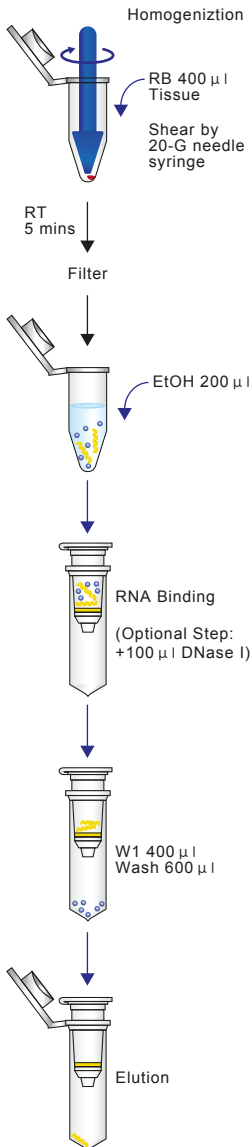
Troubleshooting

| Problem | Possible Reasons / Solution |
|----------------|--|
| Column clogged | <p>Sample overloading</p> <ul style="list-style-type: none"> Reduce sample volume or separate into multiple tubes. <p>Precipitate was formed at RNA Binding Step</p> <ul style="list-style-type: none"> Reduce the sample material. Prior to loading the column, break up precipitate in ethanol-added lysate. It is better for centrifugation at room temperature than 4°C. |
| Low yield | <p>Insufficient disruption and homogenization</p> <ul style="list-style-type: none"> Grind the sample under liquid nitrogen to a fine powder indeed. Indeed mix powder of samples with RB/PRB and β-mercaptoethanol well by vortexing. <p>Incomplete RNA Elution</p> <ul style="list-style-type: none"> Elute twice to increase yield. |
| RNA degraded | <p>Harvested samples were not immediately stabilized</p> <ul style="list-style-type: none"> Make sure that harvested samples were stabilized immediately. It is better to use fresh samples than frozen samples. <p>RNase contamination</p> <ul style="list-style-type: none"> Make sure all centrifuge tubes and tips used in RNA extraction are RNase-free. |

If the lysate mixture could not flow past membrane spin column after centrifugation, increase the centrifuge time until the lysate mixture passes completely.

Reference

(1) Vogelstein, B., and Gillespie, D. (1979) Proc. Natl. Acad. Sci. USA 76,615.



Note

Catalog

| Product Name | Package | Cat. No. |
|---|-------------------|-----------------|
| ATP™ Plasmid Mini Kit | 100/300 prep | APD100/APD300 |
| ATP™ Plasmid Midi Kit (Ultra Pure) | 25 prep | API25 |
| ATP™ Plasmid Maxi Kit (Ultra Pure) | 10/25 prep | APM10/APM25 |
| ATP™ 96-Well Plasmid Mini Kit | 4/10 plates | APD9604/APD9610 |
| ATP™ Plasmid Mini Binding Column | 50 pcs | PDC50 |
| ATP™ Plasmid Midi Resin Column | 10 pcs | PIC10 |
| ATP™ Plasmid Maxi Resin Column | 10 pcs | PMC10 |
| ATP™ Gel/PCR DNA Fragments Extraction Kit | 100/300 prep | ADF100/ADF300 |
| ATP™ 96-Well Gel/PCR DAN Extraction Kit | 4/10 plates | ADF9604/ADF9610 |
| ATP™ 96-Well SEQ Dye Clean Up Kit | 4/10 plates | ADC9604/ADC9610 |
| ATP™ Fragment DNA Binding Column | 50 pcs | DFC50 |
| ATP™ Genomic DNA Mini Kit (Blood/Culture Cell/Bacteria) | 100/300 preps | AGB100/AGB300 |
| ATP™ Genomic DNA Mini Kit (Tissue) | 50/300 preps | AGT050/AGT300 |
| ATP™ Genomic DNA Mini Kit (Plant) | 100 preps | AGP100 |
| ATP™ Genomic DNA Maxi Kit (Fresh Blood) | 25 prep | AGBM25 |
| ATP™ Genomic DNA Maxi Kit (Frozen Blood) | 25 prep | AGDM25 |
| ATP™ Plant Genomic DNA Maxi Kit | 25 prep | AGPM25 |
| ATP™ 96-Well Genomic DNA Kit | 4/10 plates | AGB9604/AGB9610 |
| ATP™ Reagent Genomic DNA Kit | For 100ml blood | AGE100 |
| ATP™ Genomic DNA Binding Column | 50 pcs | GDC50 |
| ATP™ RNA Mini Kit (Blood/Culture Cell/Bacteria) | 50 prep | ARB050 |
| ATP™ RNA Mini Kit (Tissue) | 50 prep | ART050 |
| ATP™ RNA Mini Kit (Plant) | 50 prep | ARP050 |
| ATP™ Viral Nucleic Acid Mini Kit | 50 prep | AVR050 |
| ATP™ 96-Well Viral Nucleic Acid Kit | 4/10 plates | AVR9604/AVR9610 |
| ATP™ RNA Maxi Kit | 10 prep | ARTM10 |
| ATP™ Plant RNA Maxi Kit | 10 prep | ARPM10 |
| ATP™ RNA Binding Column | 50 pcs | RBC50 |
| Proteinase K | 11 mg/kit | APK000011 |
| RNase A | 0.2 ml (50 mg/ml) | ARA500200 |
| RNase A | 1.5 ml (50 mg/ml) | ARA501500 |

